

ANDREW GOBEA:

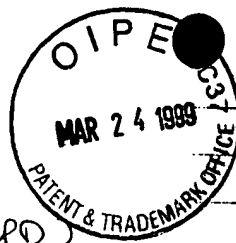


EXHIBIT
1

Received w/c cord blood (w CPD)
PPE monob 30%
PPE poly 70%

added 15 ml 1x PBS to bring to incubation volume of 45 ml.

Added 1.5 ml antibody (12.8)
incubated 25 ml.

Primed cell prep "ceprate".

spin cells. Resuspended in 1x PBS to a final volume of 300 cc in bag.

Ran through column.

Unadsorbed portion → spun down and consolidated in 1x PBS for incubation.

75 ml for incubation (added HSA)
1.5 ml antibody (12.8) 25 min. Spun down following incubation. Rsp'd to vol. of 300 cc in bag.
Ran through 2nd column.

stem cell portions from Runs 1 & 2 were combined (after counts done and samples removed for staining)

total cells 2.8×10^6 for transduction

BBMM +

123 SANDC

WON

146 SANDC

WON

SLF AUBIE

WON

final concn

will be dil

cells are

concentrated

BBMM: FB

PB

2.8×10^6 cells

Put in

BBMM + 3161SCF (for 500ml of media)

123 SANDOZ * Y0230092 stock at 150ug/ml
want final: 20ug/ml x 2 ... 20ug

133ul add

126 SANDOZ * Y0450392 stock at 150ug/ml
want final: 50ug/ml x 2 ... 50ug

333ul add

SCF AUBIEN * 1509F2 stock at 1.5mg/ml = 1500ug/ml
want final: 100ug/ml x 2 ... 100ug

167ul add

final concentrations are doubled since the media
will be diluted 1:2 w/ viral supernatant.
Cells are therefore incubated with the correct
concentrations.

BBMM: FBS Genuni lot# A26003H
BSA #115

2.8×10^6 cells want final: 5×10^4 cells.

Put in 2 T75

30ml each: 15ml B365 051193

15ml LPSU^{GT} lot# 53

+ protamine sulfate 240ul
of 1:20 diluted 50

))

rotation

to
bag.

).

in down
f 300cc in bag.

42
and

Cord Blood cells pre processing:

CPUs

SET 143

Start:

Plate #	Sample	# Cells	# ul/ml media
-G418	1ab	5×10^4	50
+G418	2ab		50
-G418	3ab	1.4×10^5	100
+G418	4ab		100

adsorber
fraction:

adsorber
fraction

CPUs Post transduction: SET 144

Plate #	# Cells	# ul
-G418	500	7
↓	1000	14
	2000	28
+G418	500	7
↓	1000	14
	2000	28

(yields)
adsorber

count:

$$\bar{x} = 34$$

$$1.2 \times 10^4 = 6.8 \times 10^5 \text{ Cl/ml}$$

$$15.5 \text{ ml} = 3.7 \times 10^6 \text{ C}$$

Transfused on 5/15/93
No transduced stem cells

G418	7ab	1000	20
-	8ab	2000	40

adsorber
yields from
fraction

Start:

 $5 \times 10^8 \text{ c}$ PRE
 0.71% Post ab
 0.22% *ul/ml media $*34+ = 3.6 \times 10^6 \text{ c}$ $= 1.1 \times 10^6 \text{ c}$ adsorbed
fraction #1: $2 \times 10^6 \text{ c}$ FL1 FL2 gate
 31.94% FL1 FL2 gate
 20.81% $*34+ = 0.64 \times 10^6 \text{ c}$ $= 0.42 \times 10^6 \text{ c}$ adsorbed
fraction #2: $0.8 \times 10^6 \text{ c}$ 2.46% 5.80% $\pm 34+c = 0.02 \times 10^6 \text{ c}$ $0.05 \times 10^6 \text{ c}$

(yields)

adsorbed #1:

PRE & FL1/FL2 gate
 $\frac{0.64 \times 10^6 \text{ c}}{3.6 \times 10^6 \text{ c}} = \boxed{17.8\%}$ PRE & FL1/FSC gate
 $\frac{0.42 \times 10^6 \text{ c}}{3.6 \times 10^6 \text{ c}} = \boxed{11.7\%}$ post ab & FL1/FL2 gate
 $\frac{0.64 \times 10^6 \text{ c}}{1.1 \times 10^6 \text{ c}} = \boxed{58.2\%}$ Post ab & FL1/FSC gate
 $\frac{0.42 \times 10^6 \text{ c}}{1.1 \times 10^6 \text{ c}} = \boxed{38.2\%}$

adsorbed #2:

~~adsorbed from~~
~~medium~~

PRE & FL1/FL2

~~adsorbed from~~

PRE & FL1/FSC

ZACHARY RIGGINS

5/14/93

Rec'd 200 cc cold Bused

PRE: $\frac{morb}{109}$ \checkmark $\frac{poly}{109}$

$$218 \times 50 = 10.9 \times 10^6 \text{ clml}$$

$$\times 200 \text{ ml} = 2.2 \times 10^9 \text{ C} \quad \text{start}$$

Added 3 vials (4.5 ml) 12.8 ab.
inc. 25 min.

spindown. Rsp'd. in 1x PBS to 300 ml
in bag.

Ran through column.

spin down unadsorbed fraction for 2nd ab
incubation.

spin stem cell fraction to Rsp'd. in
smaller volume for count.

counts:

unadsorbed

$\frac{morb}{67}$ \checkmark $\frac{poly}{102}$

$$10.9 \times 50 \times 10^6$$

$$= 8.5 \times 10^6 \text{ clml} \times 225 \text{ ml}$$

$$= 1.9 \times 10^9 \text{ C}$$

stem

$\frac{morb}{172}$ \checkmark $\frac{poly}{16}$

$$188 \times 2 \times 10^4$$

$$= 3.8 \times 10^6 \text{ clml} \times 5.5 \text{ ml}$$

$$= 20.7 \times 10^6 \text{ C}$$

inc
12.8
Spd
Rsp
Ran

con

1

0

3

3.

=

per
froz

con

26x1
won

= 2 =

13.51

LA 50

incubated unadsorbed fraction w/ 4.5 ml
 12.8 ab. for 25 min.
 spun down.

Put in 300ml in bag (w/ 1x PBS)
 Ran through 2nd column.

counts:

unadsorbed
 monos polys
 30 33

$63 \times 53 \times 10^3$

$3.15 \times 10^4 \text{ cpm} \times 60 \text{ ml}$
 $= 1.9 \times 10^9 \text{ c}$



stem
 monos polys
 58 4

$62 \times 2 \times 10^4$

$= 1.2 \times 10^6 \text{ cpm}$
 $\times 5 \text{ ml} = 6 \times 10^6 \text{ c}$

percolled/ficoll'd
 freeze \Rightarrow LWT(2)

combined stem cell fractions

$26 \times 10^6 \text{ c}$ for transduction

want final $[T] = 5 \times 10^4 \text{ cpm}$ \therefore

530 ml total

$- 2 = 260 \text{ ml supe}$

260 ml media

13 flasks 40 ml / flask

20 ml supe

20 ml media (B365)

+ 300 ml protamine sulfate

(2)

LASN supe 539 (bottles 18/19)

$\times 10^4$
 ml $\times 55 \text{ ml}$

CFUs:

5/17/45

PRE

Plate #	Sample	# Cells	# ill
1ab	(-G418) PRE + CFUs	5.5×10^4	5
2ab	↓ (+G418)	↓	5

BBMM + 31615CF:

1L3 Sander #y0230292

1L6 Sander #y0450392

SCF AMGW #150952

Took sample to micro for sterility ✓
 each day of transduction pt.
 Stat Gram stain done (negative)
 before cells were given to baby.

5/15/93 4pm 2nd transduction:

Spun cells down from each flask
 Respd in fresh media & LASN supe
 added Protamine sulfate

5/16/93 3rd transduction 330pm.
 Repeated above.

5/17

COW

POT

REX

FOOT

SG

+G4

SG

+G4

517193 cells washed 4x
 3x in 1x PBS + Pls
 last wash in RPMI (no pls)

count: 60×10^6 c

$$\bar{x} = 15 \times 10 \times 10^4 = 1.5 \times 10^6$$

$$\times 40 \text{ ml} = 60 \times 10^6 \text{ c}$$

Put in 5cc into 10cc syringe

Reinforced on 517193 (w/df)

Post-trans. CPUs: 59746

sample plate #		# cells	# ul
OG418	1ab	500	4
↓	2ab	1000	8
↓	3ab	2000	16
+G418	4ab	500	4
↓	5ab	1000	8
↓	6ab	2000	16
OG418	7ab	1000	24
+G418	8ab	1000	24

ry ✓
 wire)
 > baby
 lion
 in flock
 SN supe

> 30pm.